

Detection of IFN- γ Secretion by T Cells Collected Before and After Successful Treatment of Early Lyme Disease

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Background. Current serodiagnostics for Lyme disease lack sensitivity during early disease, and cannot determine treatment response. We evaluated an assay based on QuantiFERON technology utilizing peptide antigens derived from *Borrelia burgdorferi* to stimulate interferon-gamma (IFN- γ) release as an alternative to serodiagnosis for the laboratory detection of Lyme disease.

Methods. Blood was obtained from patients with erythema migrans before (n = 29) and 2 months after (n = 27) antibiotic therapy. IFN- γ release was measured by enzyme-linked immunosorbent assay (ELISA) following overnight stimulation of whole blood with the peptide antigens, and compared to the results of standard serological assays (C6, ELISA, and Western blot).

Results. IFN- γ release was observed in pretreatment blood of 20 of 29 (69%) patients with Lyme disease. Following antibiotic treatment, IFN- γ was significantly reduced ($P = .0002$), and was detectable in only 4 of 20 (20%) initially positive patients. By contrast, anti-C6 antibodies were detected in pretreatment sera from 17 of 29 (59%) subjects, whereas only 5 of 29 (17%) patients had positive Western blot seroreactivity. Antibody responses persisted and expanded following treatment.

Conclusions. Our findings suggest that measurement of IFN- γ after incubating blood with *Borrelia* antigens could be useful in the laboratory diagnosis of early Lyme disease. Also, after antibiotic treatment, this response appears to be short lived.

Keywords. *Borrelia burgdorferi*; IFN- γ ; Lyme disease; T cell; cytokine release assay.

The detection of antibodies to *Borrelia burgdorferi* is the standard method for the laboratory diagnosis of Lyme disease. Whether using lysates of whole *Borrelia* species, mixtures of recombinant proteins, or specific peptide antigens (eg, C6, PepC10) as assay targets [1–6], current serological assays rarely exceed a sensitivity of 50% in the positive detection of antibody in early disease. In addition, these antibody detection assays do not provide accurate information concerning treatment response, as antibody levels often remain elevated for years after the infection has been cleared [5–7]. New approaches are therefore needed to overcome the shortcomings of current serologic assays.

Antigen-specific T-cell activation is typically initiated shortly after infection. The expanding cell population secretes cytokines that, among other activities, drives the development of a mature antibody response [8–10]. Following the resolution of infection, the T-cell response wanes, which results in decreased cytokine secretion (or a shift away from proinflammatory cytokine secretion) and rapid contraction of the activated T-cell population. Therefore, a test that monitors T-cell activation might be a useful

adjunct to traditional serologic testing methods, especially because the results may provide more accurate information on the presence of active infection compared to antibody responses.

Early attempts to evaluate the utility of monitoring T-cell responses in patients with Lyme disease yielded inconclusive results [11–14]. However, these studies relied prominently on T-cell proliferation as a measurement of T-cell activity, and this approach can suffer from a significant lack of specificity [13]. Furthermore, cytokines, including interferon gamma (IFN- γ), have been shown to inhibit T-cell proliferation under certain conditions [14], which would in turn reduce the usefulness of proliferation as a marker of infection. On the other hand, antigen-induced cytokine release may be a more reliable (albeit indirect) method to confirm T-cell activation [15, 16]. Forsberg et al [15] demonstrated that detection of IFN- γ provided diagnostically relevant information for confirming neurologic Lyme disease, while Jin et al [16] reported that activated T cells from patients with Lyme disease produced IFN- γ following ex vivo stimulation with decorin binding protein A, outer surface protein C (OspC), p100, or vmp-like sequence lipoprotein E.

Despite these findings, the clinical utility of a test that measures T-cell immunity during Lyme disease has not been fully evaluated. We evaluated an assay, based on QuantiFERON technology for *Mycobacterium tuberculosis* infection, to detect IFN- γ secretion in whole blood from patients with early Lyme disease after overnight incubation with a cocktail of peptides derived from the *Borrelia* antigens p66, decorin binding protein B (DbpB), OspC, and flagellin (41 kDa). We compared the results,

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before and after appropriate antibiotic therapy, to those obtained using a standard C6 enzyme-linked immunosorbent assay (ELISA) and commercially available Lyme disease Western blot.

MATERIALS AND METHODS

Peptide Antigens

Full-length sequences of p66, DbpB, flagellin, and OspC from the B31 strain of *B. burgdorferi* sensu stricto were aligned against other *Borrelia* species using the protein basic local alignment search tool (BLASTp) on the National Center Biotechnology Information website. These antigens were selected because they are expressed in early disease. OspC is a major *B. burgdorferi* surface in early Lyme disease [4], and the other proteins are expressed constitutively [4, 7, 8] by the spirochete. In addition, preliminary findings from patients with well-documented Lyme disease generated encouraging results. A proprietary combination of 33 peptides (ranging from 15 to 25 amino acids) was generated from regions of the 4 antigens that were highly conserved among *Borrelia* species (>80% identity) and also distinct from non-*Borrelia* proteins (<50% identity). The peptides were dissolved and adjusted to a concentration of 3.3 mg/mL prior to preparing a stock cocktail that contained 100 µg/mL of each peptide in phosphate-buffered saline (pH 7.2).

Patients and Controls

Whole blood was collected from adult (≥18 years old) patients with physician-diagnosed early Lyme disease characterized by a history of tick exposure and 1 or more (≥5-cm annular lesion) erythema migrans (EM) skin lesions. As a control, blood was also obtained within 1–2 weeks after completing antibiotic treatment from patients with *Anaplasma phagocytophilum* infection (anaplasmosis), confirmed by positive polymerase chain reaction blood test at the Gundersen Health System. The subjects with early Lyme disease or anaplasmosis were each treated with 100 mg of doxycycline twice daily for a minimum of 10 days [17, 18]. Healthy adult (≥18 years old) volunteers who resided in Lyme disease foci [19, 20] in Wisconsin (n = 18) or Maryland (n = 169) and had no prior history of Lyme disease were also included as controls. Informed consent was obtained prior to enrollment, and protocols were approved by the Gundersen Health System Institutional Review Board. Each Lyme disease subject completed a questionnaire that documented epidemiology, history of tick bite, and duration and severity of clinical manifestations. Each Lyme disease subject was also queried immediately prior to the convalescent blood draw to confirm satisfactory resolution of Lyme disease–related symptoms.

Collection and Processing of Blood

Blood samples from the subjects with EM, early Lyme disease, were collected at presentation (pretreatment) and approximately 60 days after completing a course of doxycycline therapy.

Blood samples from the healthy volunteers or from patients with anaplasmosis served as controls. All blood samples were drawn into 2 lithium-heparin tubes; 1 tube was aliquoted into 1-mL volumes that were then transferred to separate sterile pyrogen-free tubes that were either empty (nil), preloaded with mitogen, or preloaded with a 10-µL volume of the peptide pool (1 µg/mL of each peptide final concentration). The tubes were then mixed thoroughly and incubated for 16–24 hours at 37°C. The remaining tube of blood was centrifuged (3000g for 15 minutes), and the separated plasma was stored at –80°C until tested.

Detection of IFN-γ

After incubation, the blood plasma from the blood collection tubes was harvested by centrifugation at 3000g for 15 minutes, and the amount of IFN-γ in the plasma was determined using the QuantiFERON ELISA (Qiagen, Germantown, Maryland) as instructed by the manufacturer. Results in the test samples were reported in international units (IU)/mL relative to a standard curve prepared by testing dilutions of a recombinant human IFN-γ standard. The nil sample was used to adjust for background reactivity caused by heterophile antibody or nonspecific IFN-γ; nil tube reactivity was subtracted from the IFN-γ value obtained using the subject's blood sample. The mitogen sample was used as a positive control to ensure the integrity of the lymphocytes in the blood sample.

C6 ELISA

The C6 peptide (CMKKDDQIAAAMVLRGMAKDGQFALK) was synthesized (>95% purity) with biotinylated amine terminal ends (GenScript, Inc, Piscataway, New Jersey), and the ELISA was performed as described previously [21]. Significant cutoff values were determined by testing archived serum samples from normal volunteers with no previous suspicion of Lyme disease (n = 30), sera that contained rheumatoid factor (n = 10) or antinuclear antibodies (n = 10), and sera from patients with Epstein-Barr virus (n = 10), cytomegalovirus (n = 10), syphilis (n = 10), Rocky Mountain spotted fever (n = 4), *Borrelia hermsii* relapsing fever (n = 14), or anaplasmosis (n = 13). The mean optical density (OD) value was calculated and OD values ≥2 standard deviations above the mean (immunoglobulin M [IgM] ≥ 0.532, immunoglobulin G [IgG] ≥ 1.206) were considered significant. The cutoff values resulted in detection of IgG antibodies in the serum from 1 patient with antinuclear antibodies and 1 with relapsing fever. Positive and normal serum controls were also included on each ELISA plate.

Western Blotting

Western blot reactivity was detected by a commercially available kit (Marblot, Trinity Biotech, Carlsbad, California) according to the manufacturer's recommendations. The criteria for serodiagnostic confirmation of Lyme disease recommended by the Centers for Disease Control and Prevention (CDC) [22] were used to determine positivity (for IgM: at least 2 of the protein bands

23, 39, or 41 detected; for IgG: at least 5 of the protein bands 18, 23, 28, 30, 39, 41, 45, 58, 66, or 93 detected).

Statistical Analyses

The Wilcoxon rank-sum test was used to examine the results, and *P* values $\leq .05$ were considered significant.

RESULTS

Study Participants

A total of 29 adult patients with physician-diagnosed Lyme disease (20 males, 9 females) were enrolled in the study (Table 1). Twenty-seven of 29 patients returned to provide a convalescent blood sample approximately 2 months after completing antibiotic therapy. The subjects had single (*n* = 23) or multiple (*n* = 6) EM lesions that met the CDC surveillance criterion (>5-cm red round lesion with or without central clearing), and 19 patients had removed an *Ixodes scapularis* (*n* = 12) or unidentified (*n* = 7) tick within the 3 weeks prior to developing the skin lesion. Two individuals had no additional clinical or constitutional complaints. The most common abnormalities in the remaining patients were headache (*n* = 23), stiff neck (*n* = 15), and myalgia

(*n* = 15). Each patient completed a minimum of 10 days of doxycycline therapy, which resulted in a complete cessation of clinical and constitutional abnormalities; no patients reported abnormal symptoms during the convalescent visit.

Detection of Anti-C6 Antibodies and Western Blot Reactivity

Acute and convalescent serum samples were tested for anti-C6 antibodies and Western blot reactivity. Anti-C6 antibodies were detected in the pretreatment sera from a total of 18 (59%) patients (Table 2), with 16 (55%) patients positive for IgM antibodies and 9 (31%) patients positive for IgG antibodies. In the 2-month posttreatment sera, anti-C6 IgM remained elevated in 11 patients and decreased to an undetectable level in 4 previously positive patients, and 1 previously negative individual seroconverted to IgM positivity. IgG anti-C6 antibody levels increased in convalescence and were detected in 14 of 27 (51.9%) of posttreatment patients. In contrast, both IgM and IgG Western blot sensitivity was very poor in both pretreatment and convalescent sera (Table 3). Although IgM and IgG Western blots detected antibodies that bound to at least 1 protein in many subjects, only 5 (17%) subjects developed antibody responses

Table 1. Epidemiology and Clinical Signs/Symptoms of Patients With Early Lyme Disease

Subject No.	Sex	Age, y	Documented (<3 wk) Tick Bite	Duration of Illness, d	Erythema Migrans Lesion(s)	Other Complaints/Abnormalities
1	M	54	Yes (deer tick)	10	Single (back)	Headache
2	F	52	No	3	Single (back)	Headache, fever, fatigue, myalgia
3	M	52	No	3	Single (thigh)	Headache, chills, fatigue, arthralgia
4	M	24	Yes (deer tick)	4	Single (thigh)	Headache, stiff neck, myalgia, arthralgia
5	M	58	Yes (deer tick)	2	Single (shoulder)	Arthralgia
6	F	62	Yes (deer tick)	3	Single (thigh)	None
7	M	30	Yes (deer tick)	5	Single (back)	Headache, fever, fatigue, stiff neck
8	M	61	Yes (deer tick)	11	Single (back)	Fatigue, myalgia
9	M	49	Yes (deer tick)	7	Single (trunk)	Headache, fever, stiff neck, fatigue, myalgia
10	F	59	Yes (deer tick)	3	Single (chest)	Headache, fever, stiff neck, myalgia
11	M	51	Yes (deer tick)	7	Single (hip)	Headache, stiff neck, myalgia
12	M	69	Yes (unidentified)	4	Single (thigh)	None
13	F	41	Yes (unidentified)	3	Single (back)	Headache, fever, fatigue, myalgia, arthralgia
14	M	74	Yes (deer tick)	2	Single (trunk)	Headache, fever, stiff neck, myalgia, arthralgia
15	F	68	No	3	Single (abdomen)	Lymphadenopathy
16	F	68	Yes (unidentified)	7	Single (abdomen)	Headache, fever, fatigue, myalgia, arthralgia
17	M	71	Yes (unidentified)	5	Single (arm)	Headache, stiff neck, fatigue, arthralgia
18	M	71	Yes (unidentified)	6	Single (back)	Headache, stiff neck, fatigue, arthralgia
19	F	48	Yes (unidentified)	3	Single (groin)	Headache, fever, fatigue
20	F	69	No	5	Single (shoulder)	Headache, stiff neck, fatigue
21	M	62	Yes (deer tick)	20	Single (back)	Headache, fever, stiff neck, arthralgia, myalgia
22	M	73	Yes (unidentified)	7	Single (shoulder)	Headache, fatigue
23	F	35	No	14	Single (back)	Headache, stiff neck, arthralgia, myalgia
24	M	59	No	4	Multiple (back)	Headache, fever, stiff neck, myalgia, arthralgia
25	M	22	No	6	Multiple (abdomen)	Headache, fever, stiff neck, myalgia, arthralgia
26	M	31	No	19	Multiple (back)	Headache, stiff neck, fatigue, myalgia
27	M	59	Yes (deer tick)	6	Multiple (trunk)	Fever, fatigue
28	M	19	No	6	Multiple (arm, torso)	Headache, fever, stiff neck, arthralgia, myalgia
29	M	36	No	30	Multiple (thigh, groin)	Headache, fever, stiff neck, arthralgia, myalgia

Table 2. Detection of Anti-C6 Antibodies in Early Lyme Disease Sera Collected Pretreatment or 2 Months After Antibiotic Therapy

Subject	Significant ^a (+/-) Levels of Anti-C6 Antibodies Detected:			
	Pretreatment		2 mo Posttreatment	
	IgM	IgG	IgM	IgG
1	+	-	-	-
2	-	-	-	-
3	+	+	+	+
4	+	+	+	+
5	-	-	-	-
6	-	-	-	-
7	+	-	-	+
8	+	-	+	+
9	+	+	+	+
10	-	-	+	-
11	+	-	-	-
12	-	-	-	-
13	+	-	-	+
14	-	-	-	+
15	-	+	-	+
16	-	-	-	-
17	-	-	-	-
18	-	-	NT	NT
19	+	-	+	+
20	+	+	+	+
21	+	+	+	+
22	-	-	-	+
23	+	-	+	-
24	-	-	-	-
25	+	-	+	-
26	+	+	+	+
27	-	-	-	-
28	+	+	NT	NT
29	+	+	+	+
Total, No. (%)	16 (55)	9 (31)	12 (44)	14 (52)

Abbreviations: IgG, immunoglobulin G; IgM, immunoglobulin M; NT, not tested.

^a Significant refers to ≥ 2 standard deviations (IgM > 0.532, IgG > 1.206) above mean optical density value of sera from normal volunteers (n = 30), sera with rheumatoid factor (n = 10) or antinuclear antibodies (n = 10), and sera from patients with Epstein-Barr virus (n = 10), cytomegalovirus (n = 10), syphilis (n = 10), Rocky Mountain spotted fever (n = 4), *Borrelia hermsii* relapsing fever (n = 14), or anaplasmosis (n = 13).

of sufficient diversity to fulfill the CDC-recommended criteria [22] (minimum 2 IgM or 5 IgG Western blot bands) for serologic confirmation of Lyme disease prior to the antibiotic therapy, and only 7 (26%) patients could be considered positive at 2 months posttreatment. In the majority of patients, antibody levels detected by either the C6 ELISA or Western blotting remained elevated or expanded despite the appropriate antibiotic therapy and resolution of clinical abnormalities.

IFN- γ Responses Before and After Antibiotic Therapy

A baseline value for IFN- γ secretion was first determined by incubating whole blood from healthy volunteers (n = 187) and patients with anaplasmosis (n = 5) (mean IFN- γ = 0.036 IU/mL

[range, 0–1.02 IU/mL]; Supplementary Table 1). A positive cut-off value was established by calculating 3 standard deviations above the mean of the control values (≥ 0.33 IU/mL). We then determined the levels of IFN- γ released after overnight culture of whole blood incubated with the cocktail of *Borrelia*-specific antigens. Blood from 20 of the 29 patients with early Lyme disease (69%) produced significant (≥ 0.33 IU/mL) levels of IFN- γ prior to treatment with antibiotics (Table 4). The magnitude of the responses varied widely among the patients (mean, 1.46 IU/mL \pm 2.26 IU/mL). Interestingly, the levels of IFN- γ did not closely correlate with either the number of EM lesions (single vs multiple) or severity of the clinical/constitutional manifestations (Tables 1 and 4). Only 6 of 192 (3.2%) control blood samples, all from healthy donors, produced significant levels of IFN- γ (mean, 0.49 IU/mL [range, 0.33–1.02 IU/mL]).

In the EM group providing a convalescent blood sample following antibiotic treatment, the IFN- γ response was decreased (n = 3) or no longer significant (n = 14) in those that were previously positive (P = .0002; Table 4 and Figure 1). The level of IFN- γ remained similar in only 1 returning patient whose pretreatment blood draw yielded a positive result, whereas levels remained undetectable in the 9 (31%) subjects who did not produce significant levels of IFN- γ prior to treatment (Table 4). Therefore, in contrast to the antibody responses detected by the C6 ELISA or Western blotting, decreased levels or inability to detect IFN- γ correlated closely with the resolution of the early Lyme disease symptoms following treatment.

DISCUSSION

Traditional antibody tests, the current mainstay of the laboratory diagnosis of Lyme disease, lack sensitivity during early illness. They are also not useful for predicting response to treatment because the antibodies typically persist long after the spirochetes are eliminated by successful antibiotic therapy [5, 6]. This is especially problematic because the long-term seropositivity and the nonspecific abnormalities (eg, arthralgia, myalgia) associated with Lyme disease have continued to cause considerable confusion concerning the success or failure of antibiotic therapy. We therefore performed a preliminary evaluation of the utility of a QuantiFERON technology-based test that quantitates the level of IFN- γ released in blood samples after incubation with *Borrelia ssp.* specific peptide antigens for confirming Lyme disease and also provides accurate prediction of successful antibiotic therapy.

Our findings showed that incubating whole blood from patients with early Lyme disease and *Borrelia* species peptides (69%) results in secretion of significant amounts of IFN- γ , thus confirming previous reports [15, 16] of IFN- γ production during infection with *B. burgdorferi*. As secretion was dependent upon incubation with the peptide antigens, we presume that CD4⁺ or CD8⁺ T cells were the primary source of the cytokine, but additional studies are required to confirm this

Table 3. Confirmation of Early Lyme Disease by Western Blotting

Subject	Western Blot Positivity ^a (+/-)			
	Pretreatment		2 mo Posttreatment	
	IgM (Bands)	IgG (Bands)	IgM (Bands)	IgG (Bands)
1	-	-	-	- (41)
2	-	-	-	- (41)
3	- (41)	- (41)	- (41)	- (41)
4	-	- (39, 41, 45)	-	- (39, 41)
5	-	-	-	- (41)
6	-	-	-	-
7	- (23)	- (41)	- (23)	- (23, 39, 41, 45)
8	-	- (41)	-	- (41, 58)
9	+ (23, 41)	- (23, 41, 45)	+ (23, 41)	- (23, 41, 45)
10	-	-	- (23)	- (23, 39, 41)
11	- (23)	-	+ (23, 41)	-
12	-	- (41)	-	- (41)
13	-	-	-	-
14	-	-	- (23)	- (23, 41)
15	- (23)	- (41, 45, 58)	- (23)	+ (18, 41, 45, 58, 66)
16	- (23)	-	- (23)	-
17	-	- (41)	-	- (41)
18	-	- (41)	NT	NT
19	- (23)	- (41)	- (23)	- (41)
20	- (23)	- (23, 39, 93)	- (23)	- (23, 39, 41, 93)
21	+ (23, 41)	- (31, 41, 45)	+ (23, 41)	- (31, 41, 45)
22	-	- (18, 23, 39, 41)	- (23)	+ (18, 23, 39, 41, 45)
23	-	-	-	- (41)
24	- (23)	- (23, 41)	+ (23, 39, 41)	- (23, 39, 41)
25	- (23)	-	+ (23, 41)	- (23, 41)
26	+ (23, 39, 41)	- (23, 39, 41)	+ (23, 39, 41)	- (23, 39, 41, 45)
27	-	-	-	-
28	+ (23, 39, 41)	+ (23, 39, 41, 45, 93)	NT	NT
29	+ (23, 39, 41)	- (23, 39, 41, 45)	- (23)	- (39, 41)
Total, No. (%)	5 (17)	1 (3)	6 (22)	2 (7)

Abbreviations: IgG, immunoglobulin G; IgM, immunoglobulin M; NT, not tested.

^a Positive (+) result based on Centers for Disease Control and Prevention criteria for Western blot positivity: IgM at least 2 of the 23, 39, or 41 bands, and IgG at least 5 of 18, 23, 28, 30, 39, 41, 45, 58, 66, or 93 bands.

hypothesis. IFN- γ secretion varied widely among patients; however, this is expected from a diverse human population. Interestingly, the levels of IFN- γ also did not correlate closely with either the magnitude of the skin infection (single or multiple EM) or the number and severity of abnormal clinical signs/symptoms. IFN- γ levels in nil tubes were similar for both patients with single and those with multiple EM (Supplementary Table 2); thus, the lack of a difference between the 2 groups is not attributable to differences in baseline IFN- γ . The significance of this is unclear; however, this study only included a small number of patients residing in the same geographic locale.

The IFN- γ detection test and C6 ELISA produced similar results in the detection of early disease, with the IFN- γ release assay being slightly more sensitive (69% vs 59%, respectively). Both assays were significantly more sensitive than Western blotting (17%) for the detection of early disease. Although the overall

rate of detection was similar for the C6 ELISA and IFN- γ release assay, analysis of individual patients revealed that the results of the 2 assays often diverged, as 11 patients were positive for only anti-C6 antibodies or only IFN- γ . A simple explanation for this divergence is that the tests used distinct target antigens. This would then also suggest that including additional antigens in the IFN- γ release assay could yield even higher sensitivity. In fact, combining the positive results from the C6 ELISA and IFN- γ release assay increases the sensitivity of confirmation of early Lyme disease in our test subjects to 83% (24 of 29).

In addition, we demonstrated that the antibody responses detected by the C6 ELISA or Western blotting typically persisted or continued to expand, even after the subjects were successfully treated with an appropriate regimen of doxycycline [17, 18]. Kowalski et al [18] showed convincingly in a large clinical study that the course of doxycycline only rarely failed to

Table 4. Detection of Interferon Gamma in Blood Samples From Patients With Early Lyme Disease Collected Pretreatment or 2 Months Posttreatment With Doxycycline

Subject	Levels (IU/mL) of IFN- γ in Blood Samples Collected ^a	
	Pretreatment	2 mo Posttreatment
1	1.16 ^b	0
2	0	0.06
3	5.03 ^b	0
4	0	0
5	0	0
6	3.86 ^b	2.61 ^b
7	1.82 ^b	0.16
8	0.45 ^b	0.02
9	1.69 ^b	1.59 ^b
10	0.78 ^b	0.46 ^b
11	0.49 ^b	0.12
12	0.29	0
13	0.33 ^b	0
14	0	0
15	2.58 ^b	0
16	2.02 ^b	0.11
17	0	0
18	10.61 ^b	1.12 ^b
19	0.42 ^b	0
20	0	0.12
21	0	0
22	2.29 ^b	NT
23	4.99 ^b	0.21
24	0.75 ^b	0.27
25	0.43 ^b	NT
26	0.47 ^b	0.20
27	0.82 ^b	0.30
28	0.66 ^b	0.05
29	0.30	0

Abbreviations: IFN- γ , interferon gamma; NT, not tested.

^a Values displayed are the IFN- γ value obtained from peptide stimulation of whole blood samples minus nil tube reactivity (Supplementary Table 2).

^b Results are ≥ 3 standard deviations above the mean value (≥ 0.33 IU/mL) of normal (n = 187) and anaplasmosis (n = 5) sera.

effectively resolve early Lyme disease. This finding therefore also confirms previous reports [6–8] documenting the inability of serology to provide accurate prediction of successful treatment. More significantly, however, we also showed that the levels of IFN- γ decreased after successful antibiotic therapy and, in fact, the response disappeared entirely in most (78%) of the patients with early Lyme disease within 2 months after the treatment had been completed. Decreases in IFN- γ were not attributable to increased baseline IFN- γ production in convalescent samples, as IFN- γ levels in both pretreatment and posttreatment nil tubes were comparable (Supplementary Table 2). Additional studies are required to more critically evaluate the ability of IFN- γ secretion to provide accurate prediction of active infection. Glickstein et al [23] reported increased levels of IFN- γ secretion by peripheral blood mononuclear cells

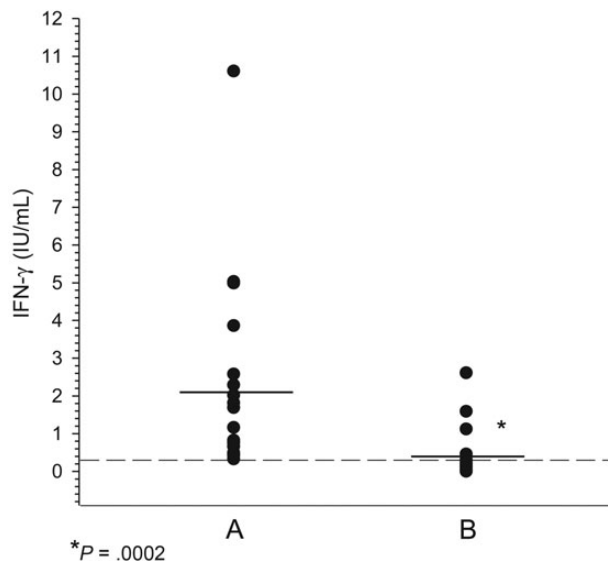


Figure 1. Detection of interferon gamma (IFN- γ) in blood samples collected approximately 2 months (column B) after treatment from patients with early Lyme disease with positive IFN- γ prior to antibiotic therapy (column A).

collected immediately after patients with early Lyme disease had completed antibiotic therapy. However, the methodologies utilized in that study were substantially different from those used here, which may contribute to the difference in observed responses. Therefore, a more comprehensive evaluation that includes earlier time points is required to confirm when the IFN- γ response begins to wane following antibiotic therapy.

Despite these promising results, several shortcomings should also be highlighted. For example, we only tested a small number of patients with Lyme disease, the patients resided in only a single geographic region, the patients were sick for days rather than weeks, and the clinical abnormalities were suggestive of only early skin infection. Moreover, we established a significant cut-off value using only normal sera from patients who resided in only 2 Lyme disease foci and a small number of subjects with anaplasmosis. Patients with other illnesses with clinical manifestations similar to those found in Lyme disease, and patients with potentially cross-reactive bacterial and viral infections should also be evaluated in future studies. In addition, the cocktail of *Borrelia*-specific peptides was chosen from only 4 proteins; other peptide antigens may be important. Therefore, additional studies to more critically define significant cutoff values, determine the effects of other peptide antigens, and evaluate subjects with localized and disseminated Lyme disease contracted from multiple geographic regions remain necessary.

In summary, the findings in this preliminary study provide strong evidence that detecting IFN- γ secretion after incubating whole blood with *Borrelia* species antigens offers considerable promise for increasing the sensitivity of laboratory diagnosis of early Lyme disease. The results also provide compelling

preliminary evidence that the cytokine response may be short lived after effective antibiotic therapy.

Supplementary Data

Supplementary materials are available at <http://cid.oxfordjournals.org>. Consisting of data provided by the author to benefit the reader, the posted materials are not copyedited and are the sole responsibility of the author, so questions or comments should be addressed to the author.

Notes

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Potential conflicts of interest. M. M. and J. B. are employed by Qiagen, Inc. S. M. C. and R. J. D. provide consulting services for Qiagen, Inc. R. J. D. and P. M. A. have relevant patents that could serve as a source for future research funding. All other authors report no potential conflicts. All authors have submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest. Conflicts that the editors consider relevant to the content of the manuscript have been disclosed.

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